

# From *N*-triisopropylsilylpyrrole to an optically active C-4 substituted pyroglutamic acid: total synthesis of penmacric acid†

Christophe Berini, Nadia Pelloux-Léon,\* Frédéric Minassian\* and Jean-Noël Denis

Received 9th June 2009, Accepted 30th July 2009

First published as an Advance Article on the web 27th August 2009

DOI: 10.1039/b911217k

The stereoselective synthesis of penmacric acid, an optically active C-4 substituted pyroglutamic acid, has been efficiently achieved through an unusual 11-step sequence starting from simple *N*-triisopropylsilylpyrrole. The key-steps are the initial addition of the pyrrole nucleus onto a chiral nitron and the obtention of the pyroglutamic acid moiety by reductive hydrogenation of the pyrrole followed by oxidation of the corresponding pyrrolidine into pyrrolidinone.

## Introduction

(*S*)-(-)-Pyroglutamic acid, also called 5-oxo-L-proline, is an intriguing optically active  $\gamma$ -lactam obtained by the thermal dehydration of L-glutamic acid (Fig. 1). The scaffold of pyroglutamic acid is present in many conformationally restrained peptides as well as peptidomimetics.<sup>1</sup> Therefore, the development of new methods for the synthesis of substituted analogues of this molecule is of great concern. Consequently, within the past decades, research efforts towards stereoselective access to this class of compounds have been made. For example, the preparation of C-4 substituted analogues often involves the use of a protected pyroglutamic acid derivative as the chiral template.<sup>2</sup> The key step consists either in stereocontrolled alkylation,<sup>3</sup> or halogenation of the lactam enolate or of the corresponding silyl enol ether.<sup>3b,4</sup> The resulting molecules are also useful intermediates in the synthesis of C-4 substituted L-glutamic acid<sup>5</sup> as well as L-proline analogues.<sup>2,6</sup> In the latter case, the reductive conversion of the pyrrolidinone into a pyrrolidine is usually achieved using boranes,<sup>7</sup> or through a two-step sequence via a hemiaminal species.<sup>8</sup>

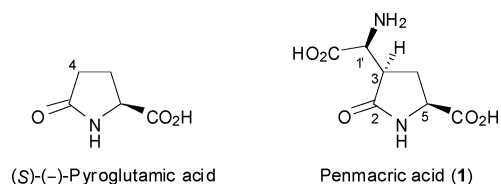


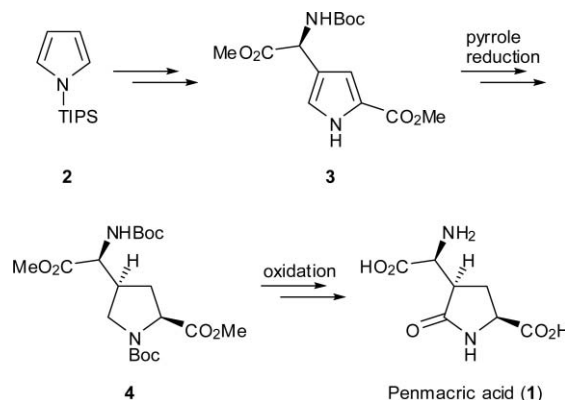
Fig. 1 Pyroglutamic frameworks.

However, the preparation of C-4 substituted chiral pyroglutamic acids from pyrrolic frameworks is not well documented despite its potential usefulness.

Following our works in the field of the total synthesis of pyrrole containing natural products,<sup>9</sup> we then decided to explore this synthetic area. Our attention was particularly focused on the

synthesis of penmacric acid (**1**) (Fig. 1). This compound was first isolated simultaneously by two independent teams from the seeds of the leguminous tree *Pentaclethra macrophylla*, also known as the “oil bean tree” that could be found in several western African countries.<sup>10,11</sup> The seeds’ extracts are both used in the local food<sup>12</sup> and as anti-inflammatories in the traditional medicines.<sup>13</sup> The structure of this compound is based on a chiral pyroglutamic acid skeleton bearing a glycine substituent at the C-4 position. Its absolute configuration was established to be (3*R*,5*S*,1'*S*) through crystallographic, spectroscopic and optical studies.<sup>14,15</sup> To date, only two research teams have described their works towards the synthesis of title compound **1**. The synthesis of this compound was first attempted by Moloney and co-workers in 2003, but the authors were not able to reach the target molecule **1**.<sup>16</sup> In 2007, Naito and co-workers described the first total synthesis of **1** in 12 steps (5.4% overall yield) as well as the synthesis of the C-1' epimer (3.0% overall yield).<sup>17</sup> None of these strategies involved the use of a functionalized pyrrole backbone as the key intermediate.

Our present strategy towards penmacric acid (**1**) implies first some selective transformations of the *N*-triisopropylsilylpyrrole **2** into the appropriate 2,4-disubstituted pyrrole **3** bearing a stereogenic center on the C-4 position. Then, a sequence involving the reduction of the pyrrole ring<sup>18</sup> followed by the oxidation of the chiral pyrrolidine **4** would give access to the target molecule **1** (Scheme 1).



Scheme 1 Proposed strategy.

Département de Chimie Moléculaire (SERCO), UMR-5250, ICMG FR-2607, Université Joseph Fourier, CNRS, BP-53, 38041 Grenoble cedex 9, France. E-mail: Frederic.Minassian@ujf-grenoble.fr; Fax: +33-4-7663-5983; Tel: +33-4-7651-4908

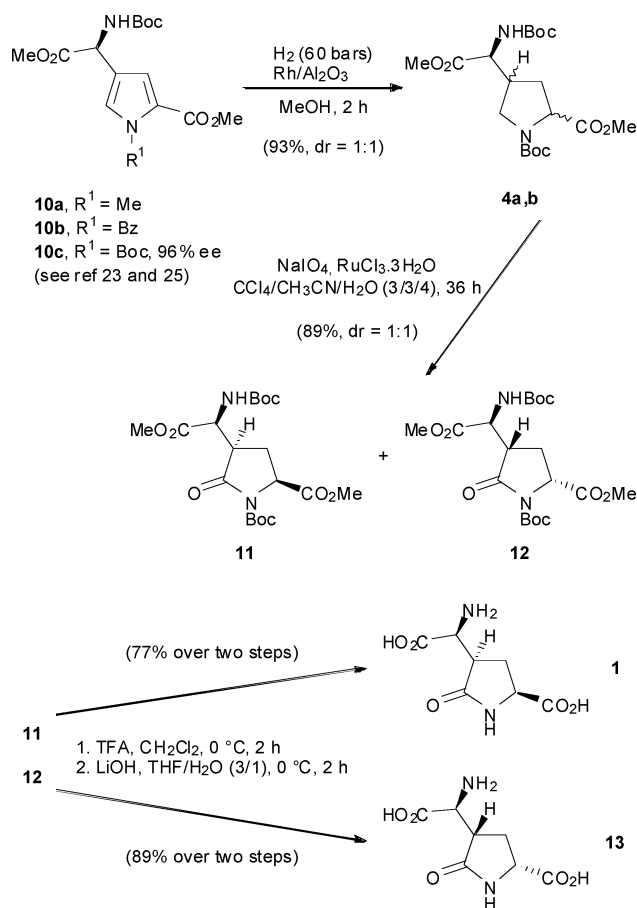
† Electronic supplementary information (ESI) available: Experimental details, copies of <sup>1</sup>H and <sup>13</sup>C NMR spectra. See DOI: 10.1039/b911217k

## Results and discussion

The introduction of the chiral glycyl substituent was first performed according to our previous works,<sup>19</sup> by addition of *N*-triisopropylsilylpyrrole **2**<sup>20</sup> onto the known cyclic chiral nitron (*S*)-**5**<sup>21</sup> under mild acidic conditions. The final methanolysis produced the C-3 pyrrolic hydroxylamine **6** as a unique regio- and diastereoisomer in 66% isolated yield (Scheme 2). Removal of the chiral auxiliary and N–O bond cleavage using palladium black as the catalyst in the presence of formic acid gave amino ester **7**.<sup>22</sup> In order to prevent any self condensation of compound **7**, protection of the amino group was performed under classical conditions to give the *tert*-butyl carbamate **8**. As selective functionalization at the  $\alpha$ -position of the pyrrole was needed in the next steps, the bulky silyl group on the nitrogen atom was first removed using TBAF in the presence of acetic acid. Subsequent trichloroacetylation of compound **9** followed by treatment of the crude mixture with sodium methoxide gave the corresponding methyl ester **3** in 90% yield as a unique regioisomer that possesses the framework of the target molecule **1**. The structure of compound **3** has been confirmed through some 1D and 2D NMR experiments.

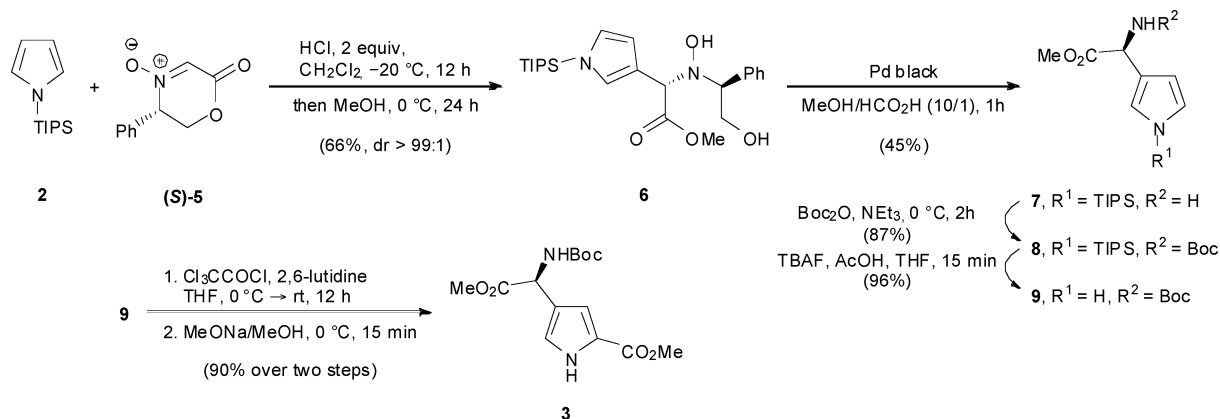
In order to explore the stereoselective hydrogenation of the pyrrole nucleus into the corresponding pyrrolidine, the protected derivatives **10a**, **10b** and **10c** were then prepared with good to excellent yields following typical procedures.<sup>23</sup> Indeed, all our preliminary attempts at reduction of the unprotected pyrrole **3**—using either Rh/Al<sub>2</sub>O<sub>3</sub> or Pt/C as the catalyst under levels of hydrogen pressure ranging from 1 to 60 bars in methanol or acetic acid—only left the unreacted starting material. Although Frontier and Jiang found that the hydrogenation of *N*-methyl pyrroles was an efficient process,<sup>18</sup> in our case compound **10a** was not reactive at all, even under a 60 bar hydrogen pressure using a catalytic amount of Rh/Al<sub>2</sub>O<sub>3</sub> in methanol. The use of the *N*-benzoyl derivative **10b** was unsuccessful as well. In this case, the non-quantitative cleavage of the benzoyl group was observed, as was previously described when using Raney nickel.<sup>24</sup>

Finally, hydrogenation of the *N*-Boc derivative **10c**<sup>25</sup> under the same conditions provided a (1:1) mixture of the pyrrolidines **4a,b** in very good yield (Scheme 3). At this stage, the two diastereoisomers were inseparable. The subsequent oxidation using ruthenium tetroxide in a ternary solvent system was then performed starting



**Scheme 3** Synthesis of title compound **1** and its stereoisomer **13**.

from this mixture.<sup>26</sup> The corresponding diastereomeric pyrrolidones **11** and **12** were obtained in 89% yield and were separated by silica gel chromatography. Careful analysis of the NMR spectra allowed the assignment of the *cis-syn* configuration for compound **11**, and *cis-anti* for compound **12** (Scheme 3). The final deprotection sequence was performed in two steps on each compound with good overall yields.<sup>27</sup>



**Scheme 2** Synthesis of compound **3**.

## Conclusions

In conclusion, our synthetic sample of penmacric acid (**1**) was obtained by an 11-step sequence with a good overall yield (6.9%) and displayed the same physical properties as the natural compound ( $[\alpha]_{\text{D}}^{20} +33.6$  ( $c$  0.070, 0.1 N HCl); lit.<sup>10</sup>  $[\alpha]_{\text{D}}^{20} +35$  ( $c$  0.070, 0.1 N HCl); consistent with an ee value of 96%). Furthermore, this original strategy allowed access to another stereoisomer **13**, which is the antipode of the known epipenmacric acid,<sup>17</sup> in 11 steps with a very good overall yield (8.0%). Further studies in the field of the synthesis of C-4 substituted pyroglutamic acid derivatives are currently being undertaken by our team.

## Experimental

### General experimental

All reactions were carried out using oven-dried glassware under an argon atmosphere. Solvents were purified prior to use by conventional methods. All other reagent-grade chemicals were used as supplied (analytical or HPLC grade) without prior purification. Thin layer chromatography was performed on aluminium plates coated with 60 PF254 silica. Plates were visualised using UV light (254 nm), followed by heating after treatment with an appropriate revelatory (KMnO<sub>4</sub>, TTC, phosphomolybdic acid, ninhydrin). Flash column chromatography was performed on Kieselgel 60 silica (40-60 mesh).

Elemental analyses were recorded by the microanalysis service of the Département de Chimie Moléculaire, Grenoble, France. Melting points were recorded on a Büchi B35 apparatus and are uncorrected. Optical rotations were recorded on a Perkin-Elmer 341 polarimeter with a water-jacketed 10 cm cell. Specific rotations are reported in 10<sup>-1</sup> deg cm<sup>2</sup> g<sup>-1</sup> and concentrations in g per 100 mL. Circular dichroisms were recorded on a JASCO J-810 spectropolarimeter. IR spectra were recorded on a Nicolet Impact-400 Fourier transform infrared spectrometer (FTIR) as either thin films on NaCl plates (thin film) or as a KBr disc (KBr disc), as stated. Selected characteristic bands are reported in cm<sup>-1</sup>. NMR spectra were recorded either on a Bruker Advance300 or on an Advance400 spectrometer in the deuterated solvent as stated. The field was locked by external referencing to the relevant deuterium resonance. Low Resolution Mass Spectra (LRMS) were recorded on a Bruker Esquire 3000 plus (ESI) or a ThermoFinnigan PolarisQ ion-trap spectrometer, using DCI (ammonia/isobutane 63/37). Accurate mass measurements were run in the "Structure et Fonction de Molécules Bioactives" laboratory, Paris, France.

**(S)-[Hydroxy-((S)-2-hydroxy-1-phenylethyl)amino]-(1-triisopropylsilylanyl-1H-pyrrol-3-yl)acetic acid methyl ester (6).** To a stirred solution of nitrone (**S**)-**5** (0.96 g, 5.02 mmol) in anhydrous CH<sub>2</sub>Cl<sub>2</sub> (25 mL) at -40 °C was added HCl (5.0 mL, 2.0 N in Et<sub>2</sub>O, 10.0 mmol) and *N*-TIPS-pyrrole **2** (1.23 g, 5.51 mmol). The resulting mixture was slowly warmed to -20 °C and then stirred at this temperature for 12 hours whereupon anhydrous MeOH (25 mL) was added. It was then allowed to react at 0 °C for 24 hours. The mixture was then treated with a saturated aqueous NaHCO<sub>3</sub> solution until pH 8–9. The aqueous layer was extracted three times with CH<sub>2</sub>Cl<sub>2</sub>. The combined organic layers were washed with brine, dried over anhydrous MgSO<sub>4</sub> and concentrated *in vacuo*. The crude product was purified by flash chromatography

on silica gel (eluent: pentane/EtOAc, 9/1 to 7/3) to afford pure *N*-hydroxylamine **6** (1.48 g, 66%) as a pale pink solid; mp 50–51 °C;  $[\alpha]_{\text{D}}^{20} +119.3$  ( $c$  1.00 in CHCl<sub>3</sub>);  $\nu_{\text{MAX}}/\text{cm}^{-1}$  (KBr disc) 3550–3350 (broad, OH  $\times$  2), 1752 (CO);  $\delta_{\text{H}}$  (400 MHz; CDCl<sub>3</sub>; Me<sub>4</sub>Si) 1.09–1.13 (18H, m, ((CH<sub>3</sub>)<sub>2</sub>CH)<sub>3</sub>Si), 1.38–1.50 (3H, m, ((CH<sub>3</sub>)<sub>2</sub>CH)<sub>3</sub>Si), 3.54–3.58 (1H, m, C(2')H<sub>2</sub>), 3.63 (3H, s, OCH<sub>3</sub>), 4.00 (1H, dd,  $J$  = 9.7 and 3.8 Hz, C(1')H), 4.23 (1H, br s, OH), 4.33–4.38 (1H, m, C(2')H<sub>2</sub>), 4.39 (1H, s, C(2)H), 6.42 (1H, dd,  $J$  = 2.4 and 1.2 Hz, pyrrolH), 6.57 (1H, s, OH), 6.70 (1H, s, pyrrolH), 6.76 (1H, t,  $J$  = 2.4 Hz, pyrrolH), 7.30–7.32 (5H, m, ArH);  $\delta_{\text{C}}$  (75 MHz; CDCl<sub>3</sub>; Me<sub>4</sub>Si) 11.7 (((CH<sub>3</sub>)<sub>2</sub>CH)<sub>3</sub>Si), 17.8 (((CH<sub>3</sub>)<sub>2</sub>CH)<sub>3</sub>Si), 52.2 (OCH<sub>3</sub>), 63.4 (C(2')H<sub>2</sub>), 66.6 (C(2)H), 67.6 (C(1')H), 110.5 (pyrrolCH), 117.8 (pyrrolC), 124.5 (pyrrolCH), 124.9 (pyrrolCH), 127.8 (ArCH), 127.9 (ArCH), 130.2 (ArCH), 140.2 (ArC), 174.0 (C=O);  $m/z$  (DCI, NH<sub>3</sub>-isobutane) 447 ([M+H]<sup>+</sup>), 431, 399, 294 (100%); Elemental analysis, calcd (%) for C<sub>24</sub>H<sub>38</sub>N<sub>2</sub>O<sub>4</sub>Si: C, 64.54; H, 8.58; N, 6.28. Found: C, 64.36; H, 8.73; N, 6.38.

**4-[(S)-tert-Butoxycarbonylamino-methoxycarbonyl-methyl]-1H-pyrrole-2-carboxylic acid methyl ester 3.** To a stirred solution of compound **9** (1.03 g, 4.05 mmol) in anhydrous THF (32 mL) at 0 °C was added 2,6-lutidine (1.20 mL, 10.3 mmol). Freshly distilled trichloroacetyl chloride (1.20 mL, 10.3 mmol) was added dropwise and the mixture was allowed to warm to room temperature. After being stirred for 12 hours, the reaction mixture was quenched with water. The aqueous layer was extracted three times with CH<sub>2</sub>Cl<sub>2</sub>. The combined organic layers were washed with saturated aqueous NaHCO<sub>3</sub> solution, brine, dried over anhydrous MgSO<sub>4</sub> and concentrated under reduced pressure. The resulting crude product was dissolved in anhydrous MeOH (81 mL) and cooled to 0 °C. Sodium methoxide (1.00 g, 18.5 mmol) was added and the mixture was stirred at 0 °C for 15 minutes. It was then acidified with 3.0 N aqueous HCl to pH 3–4. After removal of the solvent under reduced pressure, the residue was dissolved in CH<sub>2</sub>Cl<sub>2</sub>. Water was added and the aqueous layer was extracted three times with CH<sub>2</sub>Cl<sub>2</sub>. The combined organic layers were washed with brine, dried over anhydrous MgSO<sub>4</sub> and concentrated. The crude product was purified by flash chromatography on silica gel (eluent: pentane/EtOAc, 9/1 to 7/3) to afford **3** (1.14 g, 90% over two steps) as a white solid; mp 40–41 °C;  $[\alpha]_{\text{D}}^{20} +86.8$  ( $c$  1.00 in CHCl<sub>3</sub>);  $\nu_{\text{MAX}}/\text{cm}^{-1}$  (KBr disc) 3400–3200 (broad, NH), 1735 (CO), 1693 (CO);  $\delta_{\text{H}}$  (400 MHz; CDCl<sub>3</sub>; Me<sub>4</sub>Si) 1.44 (9H, s, C(CH<sub>3</sub>)<sub>3</sub>), 3.76 (3H, s, OCH<sub>3</sub>), 3.84 (3H, s, OCH<sub>3</sub>), 5.11–5.33 (2H, m, C(2)H and NH), 6.86–6.87 (1H, m, pyrrolH), 6.96–6.97 (1H, m, pyrrolH), 9.07 (1H, br s, NH);  $\delta_{\text{C}}$  (75 MHz; CDCl<sub>3</sub>; Me<sub>4</sub>Si) 28.2 (C(CH<sub>3</sub>)<sub>3</sub>), 51.3 (CH), 51.5 (OCH<sub>3</sub>), 52.5 (OCH<sub>3</sub>), 80.1 (C(CH<sub>3</sub>)<sub>3</sub>), 113.4 (pyrrolCH), 121.3 (pyrrolC), 121.4 (pyrrolCH), 123.0 (pyrrolC), 155.0 (C=O), 161.4 (C=O), 171.2 (C=O);  $m/z$  (DCI, NH<sub>3</sub>/isobutane) 313 ([M+H]<sup>+</sup>), 274, 213, 196 (100%); Elemental analysis, calcd (%) for C<sub>14</sub>H<sub>20</sub>N<sub>2</sub>O<sub>6</sub>: C, 53.84; H, 6.46; N, 8.97. Found: C, 53.95; H, 6.73; N, 8.98.

**(2S,4R)-4-[(S)-tert-Butoxycarbonylamino-methoxycarbonyl-methyl]-5-oxo-pyrrolidine-1,2-dicarboxylic acid 1-tert-butyl ester 2-methyl ester 11 and (2R,4S)-4-[(S)-tert-Butoxycarbonylamino-methoxycarbonyl-methyl]-5-oxo-pyrrolidine-1,2-dicarboxylic acid 1-tert-butyl ester 2-methyl ester 12.** To a stirred solution of pyrrolidines **4a,b** (500 mg, 1.20 mmol) in a mixture of CCl<sub>4</sub>, CH<sub>3</sub>CN and H<sub>2</sub>O (3/3/4, 24 mL) at 0 °C were sequentially added

NaIO<sub>4</sub> (1.30 g, 6.06 mmol) and RuCl<sub>3</sub>·3H<sub>2</sub>O (50 mg, 0.24 mmol). The resulting mixture was slowly warmed to room temperature and then stirred at this temperature for 36 hours. The aqueous layer was extracted three times with CH<sub>2</sub>Cl<sub>2</sub>. The combined organic layers were dried over anhydrous MgSO<sub>4</sub>, filtered through a celite pad and concentrated under reduced pressure. The crude product was purified by flash chromatography on silica gel (eluent: pentane/EtOAc, 9/1 to 1/1) to afford pyrrolidinone **11** (*cis-syn*) (230 mg, 44%) as a white solid and pyrrolidinone **12** (*cis-anti*) (233 mg, 45%) as a white solid. **Data for compound 11** (*cis-syn*): mp 69–70 °C; [α]<sub>D</sub><sup>20</sup> +15.1 (*c* 1.00 in CHCl<sub>3</sub>); ν<sub>MAX</sub>/cm<sup>-1</sup> (KBr disc) 3500–3300 (broad, NH), 1798 (CO), 1748 (CO), 1714 (CO); δ<sub>H</sub> (300 MHz; CDCl<sub>3</sub>; Me<sub>4</sub>Si) 1.44 (9H, s, C(CH<sub>3</sub>)<sub>3</sub>), 1.46 (9H, s, C(CH<sub>3</sub>)<sub>3</sub>), 1.93 (1H, ddd, *J* = 13.3, 10.9 and 8.9 Hz, 1H of C(3)H<sub>2</sub>), 2.55 (1H, ddd, *J* = 13.2, 9.7 and 8.2 Hz, 1H of C(3)H<sub>2</sub>), 3.48 (1H, dt, *J* = 10.8, and 3.0 Hz, C(4)H), 3.76 (3H, s, OCH<sub>3</sub>), 3.80 (3H, s, OCH<sub>3</sub>), 4.50 (1H, dd, *J* = 10.7 and 8.5 Hz, C(2)H), 4.53 (1H, d, *J* = 8.9 Hz, C(1')H), 5.29 (1H, d, *J* = 8.7 Hz, NH); δ<sub>C</sub> (100 MHz; CDCl<sub>3</sub>; Me<sub>4</sub>Si) 24.3 (C(3)H<sub>2</sub>), 27.7 (C(CH<sub>3</sub>)<sub>3</sub>), 28.1 (C(CH<sub>3</sub>)<sub>3</sub>), 45.7 (C(4)H), 52.4 (2 × OCH<sub>3</sub>), 52.8 (C(1')H), 57.1 (C(2)H), 80.3 (C(CH<sub>3</sub>)<sub>3</sub>), 84.0 (C(CH<sub>3</sub>)<sub>3</sub>), 148.7 (C=O), 156.1 (C=O), 170.3 (C=O), 171.1 (C=O), 171.8 (C=O); *m/z* (DCI, NH<sub>3</sub>/isobutane) 448 ([M+NH<sub>4</sub>]<sup>+</sup>), 376, 331, 231 (100%); Elemental analysis, calcd (%) for C<sub>19</sub>H<sub>30</sub>N<sub>2</sub>O<sub>9</sub>: C, 53.02; H, 7.03; N, 6.51. Found: C, 52.67; H, 7.18; N, 6.34; **Data for compound 12** (*cis-anti*): mp 50–51 °C; [α]<sub>D</sub><sup>20</sup> +14.1 (*c* 1.00 in CHCl<sub>3</sub>); ν<sub>MAX</sub>/cm<sup>-1</sup> (KBr disc) 3500–3300 (broad, NH), 1790 (CO), 1752 (broad, CO), 1718 (CO); δ<sub>H</sub> (300 MHz; CDCl<sub>3</sub>; Me<sub>4</sub>Si) 1.44 (9H, s, C(CH<sub>3</sub>)<sub>3</sub>), 1.49 (9H, s, C(CH<sub>3</sub>)<sub>3</sub>), 2.06–2.16 (1H, m, 1H of C(3)H<sub>2</sub>), 2.49–2.60 (1H, m, 1H of C(3)H<sub>2</sub>), 3.07 (1H, dt, *J* = 9.9 and 4.1 Hz, C(4)H), 3.76 (3H, s, OCH<sub>3</sub>), 3.80 (3H, s, OCH<sub>3</sub>), 4.52 (1H, dd, *J* = 8.8 and 7.7 Hz, C(2)H), 4.57 (1H, dd, *J* = 8.7 and 4.2 Hz, C(1')H), 5.64 (1H, br s, NH); δ<sub>C</sub> (100 MHz; CDCl<sub>3</sub>; Me<sub>4</sub>Si) 24.0 (C(3)H<sub>2</sub>), 27.8 (C(CH<sub>3</sub>)<sub>3</sub>), 28.1 (C(CH<sub>3</sub>)<sub>3</sub>), 45.4 (C(4)H), 52.5 (OCH<sub>3</sub>), 52.6 (OCH<sub>3</sub>), 52.9 (C(1')H), 57.0 (C(2)H), 80.3 (C(CH<sub>3</sub>)<sub>3</sub>), 84.0 (C(CH<sub>3</sub>)<sub>3</sub>), 148.9 (C=O), 155.2 (C=O), 170.5 (C=O), 171.3 (C=O), 172.1 (C=O); *m/z* (DCI, NH<sub>3</sub>/isobutane) 448 ([M+NH<sub>4</sub>]<sup>+</sup>), 376, 331, 231 (100%); Elemental analysis, calcd (%) for C<sub>19</sub>H<sub>30</sub>N<sub>2</sub>O<sub>9</sub>: C, 53.02; H, 7.03; N, 6.51. Found: C, 52.73; H, 7.09; N, 6.23.

**Penmacric acid 1.** To a stirred solution of pyrrolidinone **11** (186 mg, 0.43 mmol) in anhydrous CH<sub>2</sub>Cl<sub>2</sub> (4.3 mL) at 0 °C was added anhydrous trifluoroacetic acid (640 μL, 8.64 mmol). After being stirred for 2 hours at this temperature, solvents were evaporated under reduced pressure. The resulting crude product was dissolved in a THF/H<sub>2</sub>O mixture (3/1, 21 mL) at 0 °C. Lithium hydroxide monohydrate (72 mg, 1.72 mmol) was added and the resulting mixture was stirred for 2 hours at this temperature. It was then acidified with 0.2 N aqueous HCl to pH 3. After removal of the solvent *in vacuo*, the residue was passed through a column of Dowex 50W-X8 ion exchange resin (H<sup>+</sup> form), eluted with water and then 0.5 N aqueous ammonia. Lyophilization afforded penmacric acid **1** (67 mg, 77% over two steps) as a white hygroscopic solid: mp 194–195 °C; [α]<sub>D</sub><sup>20</sup> +33.6 (*c* 0.070 in 0.1 N HCl), lit.<sup>10</sup> [α]<sub>D</sub><sup>20</sup> +35 (*c* 0.070 in 0.1 N HCl); ν<sub>MAX</sub>/cm<sup>-1</sup> (KBr disc) 3600–2800 (broad, NH, NH<sub>2</sub> and OH), 1689 (broad, CO); δ<sub>H</sub> (300 MHz; D<sub>2</sub>O) 1.96 (1H, dd, *J* = 22.2 and 10.8 Hz, 1H of C(3)H<sub>2</sub>), 2.75 (1H, ddd, *J* = 13.1, 8.5 and 8.4 Hz,

1H of C(3)H<sub>2</sub>), 3.10 (1H, dd, *J* = 18.1 and 8.4 Hz, C(4)H), 3.94 (1H, d, *J* = 7.3 Hz, C(1')H), 4.21 (1H, t, *J* = 8.3 Hz, C(2)H); δ<sub>C</sub> (100 MHz; D<sub>2</sub>O) 29.3 (C(3)H<sub>2</sub>), 41.7 (C(4)H), 55.0 (C(1')H), 56.1 (C(2)H), 172.0 (C=O), 177.7 (C=O), 178.9 (C=O); *m/z* (ESI<sup>+</sup>) 225 ([M+Na]<sup>+</sup>), 203 ([M+H]<sup>+</sup>); HRMS (ESI<sup>+</sup>) C<sub>7</sub>H<sub>11</sub>N<sub>2</sub>O<sub>5</sub> ([M+H]<sup>+</sup>) requires 203.06625 found 203.06563.

**(2R,4S)-4-[(S)-Amino-carboxy-methyl]-5-oxo-pyrrolidine-2-carboxylic acid (13).** The same procedure applied to pyrrolidinone **12** (220 mg, 0.51 mmol) afforded penmacric acid's stereoisomer **13** (91 mg, 89% over two steps) as a white hygroscopic solid: mp 137–138 °C (decomp.); [α]<sub>D</sub><sup>20</sup> +32.4 (*c* 0.50 in 0.1 N HCl); ν<sub>MAX</sub>/cm<sup>-1</sup> (KBr disc) 3600–2800 (broad, NH, NH<sub>2</sub> and OH), 1680 (broad, CO); δ<sub>H</sub> (300 MHz; D<sub>2</sub>O) 1.93 (1H, ddd, *J* = 13.5, 10.0 and 8.4 Hz, 1H of C(3)H<sub>2</sub>), 2.65 (1H, ddd, *J* = 14.1, 11.4 and 8.8 Hz, 1H of C(3)H<sub>2</sub>), 3.47 (1H, dt, *J* = 10.0 and 3.3 Hz, C(4)H), 4.19 (1H, d, *J* = 3.0 Hz, C(1')H), 4.24 (1H, t, *J* = 8.1 Hz, C(2)H); δ<sub>C</sub> (100 MHz; D<sub>2</sub>O) 25.9 (C(3)H<sub>2</sub>), 42.7 (C(4)H), 52.8 (C(1')H), 56.0 (C(2)H), 172.6 (C=O), 177.2 (C=O), 179.2 (C=O); *m/z* (ESI<sup>+</sup>) 225 ([M+Na]<sup>+</sup>), 203 ([M+H]<sup>+</sup>); HRMS (ESI<sup>+</sup>) C<sub>7</sub>H<sub>11</sub>N<sub>2</sub>O<sub>5</sub> ([M+H]<sup>+</sup>) requires 203.06625 found 203.06561.

## Acknowledgements

We thank Dr Andrew E. Greene for his interest in our work. Financial support from the CNRS and the Université Joseph Fourier (UMR 5250, FR-2607) and a fellowship award (to C. B.) from Cluster “Chimie Durable et Chimie pour la Santé” (Région Rhône-Alpes) are gratefully acknowledged.

## Notes and references

- (a) P. K. Mandal, K. K. Kaluarachi, D. Ogrin, S. G. Bott and J. S. McMurray, *J. Org. Chem.*, 2005, **70**, 10128–10131; (b) S. Hanessian and L. Auzzas, *Acc. Chem. Res.*, 2008, **41**, 1241–1251.
- For a review, see: C. Nájera and M. Yus, *Tetrahedron: Asymmetry*, 1999, **10**, 2245–2303.
- (a) J. K. Thottathil, J. L. Moniot, R. H. Mueller, M. K. Y. Wong and T. P. Kissick, *J. Org. Chem.*, 1986, **51**, 3140–3143; (b) M. J. Beard, J. H. Bailey, D. T. Cherry, M. G. Moloney, S. B. Shim, K. A. Statham, M. J. Bamford and R. B. Lamont, *Tetrahedron*, 1996, **52**, 3719–3740; (c) J.-D. Charrier, J. E. S. Duffy, P. B. Hitchcock and D. W. Young, *Tetrahedron Lett.*, 1998, **39**, 2199–2202; (d) D. K. Dikshit and A. Maheshwari, *Tetrahedron Lett.*, 1999, **40**, 4411–4412; (e) E. L. Bentz, R. Goswami, M. G. Moloney and S. M. Westaway, *Org. Biomol. Chem.*, 2005, **3**, 2872–2882; (f) T. J. Hill, P. Kocis and M. G. Moloney, *Tetrahedron Lett.*, 2006, **47**, 1461–1463; (g) M. Volgraf, P. Gorostiza, S. Szobota, M. R. Helix, E. Y. Isacoff and D. Trauner, *J. Am. Chem. Soc.*, 2007, **129**, 260–261.
- (a) D. W. Konas and J. K. Coward, *J. Org. Chem.*, 2001, **66**, 8831–8842; (b) J. L. Cohen and A. R. Chamberlin, *J. Org. Chem.*, 2007, **72**, 9240–9247.
- M. G. Moloney, *Nat. Prod. Rep.*, 2002, **19**, 597–616 and references cited therein.
- X. Chen, D.-M. Du and W.-T. Hua, *Tetrahedron: Asymmetry*, 2002, **13**, 43–46.
- (a) M. Rodriguez, S. Terracciano, E. Cini, G. Settembrini, I. Bruno, G. Bifulco, M. Taddei and L. Gomez-Paloma, *Angew. Chem., Int. Ed.*, 2006, **45**, 423–427; (b) F. Lenda, F. Guenoun, J. Martinez and F. Lamaty, *Tetrahedron Lett.*, 2007, **48**, 805–808.
- (a) J. Ezquerra, C. Pedregal, B. Yruretagoyena, A. Rubio, M. C. Carreño, A. Escribano and J. L. García-Ruano, *J. Org. Chem.*, 1995, **60**, 2925–2930; (b) W. Xie, B. Zou, D. Pei and D. Ma, *Org. Lett.*, 2005, **7**, 2775–2777; (c) V. K.-Y. Lo, M.-K. Wong and C.-M. Che, *Org. Lett.*, 2008, **10**, 517–519.
- (a) B. Sayah, N. Pelloux-Léon and Y. Vallée, *J. Org. Chem.*, 2000, **65**, 2824–2826; (b) B. Sayah, N. Pelloux-Léon, A. Milet, J.

- Pardillos-Guindet and Y. Vallée, *J. Org. Chem.*, 2001, **66**, 2522–2525; (c) J. Patel, N. Pelloux-Léon, F. Minassian and Y. Vallée, *J. Org. Chem.*, 2005, **70**, 9081–9084; (d) J. Patel, N. Pelloux-Léon, F. Minassian and Y. Vallée, *Tetrahedron Lett.*, 2006, **47**, 5561–5563.
- 10 A. Welter, J. Jadot, G. Dardenne, M. Marlier and J. Casimir, *Phytochemistry*, 1975, **14**, 1347–1350.
- 11 E. I. Mbadiwe, *Phytochemistry*, 1975, **14**, 1351–1354.
- 12 M. O. Isichei and S. C. Achinewhu, *Food Chem.*, 1988, **30**, 83–92.
- 13 P. A. Akah and A. I. Nwambie, *J. Ethnopharmacol.*, 1994, **42**, 179–182.
- 14 L. Dupont, O. Dideberg and A. Welter, *Acta Crystallogr., Sect. B: Struct. Crystallogr. Cryst. Chem.*, 1975, **31**, 1018–1022. As it could be seen in Fig. 1, an unusual numbering scheme has been used by the authors for penmacric acid (**1**). In this paper, we prefer to use the usual pyroglutamic acid numbering scheme in place of this unusual numbering scheme.
- 15 (a) A. Welter, M. Marlier and G. Dardenne, *Bull. Soc. Chim. Belg.*, 1975, **84**, 243–252; (b) A. Welter, J. Jadot, G. Dardenne, M. Marlier and J. Casimir, *Bull. Soc. Chim. Belg.*, 1975, **84**, 453–458; (c) A. Welter, *Bull. Soc. Chim. Belg.*, 1975, **84**, 717–720.
- 16 M. Anwar, J. H. Bailey, L. C. Dickinson, H. J. Edwards, R. Goswami and M. G. Moloney, *Org. Biomol. Chem.*, 2003, **1**, 2364–2376 and references cited therein.
- 17 M. Ueda, A. Ono, D. Nakao, O. Miyata and T. Naito, *Tetrahedron Lett.*, 2007, **48**, 841–844.
- 18 C. Jiang and A. J. Frontier, *Org. Lett.*, 2007, **9**, 4939–4942.
- 19 (a) C. Berini, F. Minassian, N. Pelloux-Léon and Y. Vallée, *Tetrahedron Lett.*, 2005, **46**, 8653–8656; (b) C. Berini, F. Minassian, N. Pelloux-Léon, J.-N. Denis, Y. Vallée and C. Philouze, *Org. Biomol. Chem.*, 2008, **6**, 2574–2586.
- 20 B. L. Bray, P. H. Mathies, R. Naef, D. R. Solas, T. T. Tidwell, D. R. Artis and J. M. Muchowski, *J. Org. Chem.*, 1990, **55**, 6317–6328.
- 21 O. Tamura, K. Gotanda, J. Yoshino, Y. Morita, R. Terashima, M. Kikuchi, T. Miyawaki, N. Mita, M. Yamashita, H. Ishibashi and M. Sakamoto, *J. Org. Chem.*, 2000, **65**, 8544–8551.
- 22 These conditions gave the best results regarding the optical activity of the amino ester **7**. Using Pearlman's catalyst under a hydrogen atmosphere the reaction was nearly quantitative, but the ee was lowered to 86%. Under several other conditions, only poor ee's were observed.
- 23 (a) **10a** (prepared under a racemic form): D. G. Hulcoop and M. Lautens, *Org. Lett.*, 2007, **9**, 1761–1764; (b) **10b** (prepared under a racemic form): C. D'Silva and R. Iqbal, *Synthesis*, 1996, 457–458; (c) **10c**: J. A. Ganske, R. K. Pandey, M. J. Postich and K. M. Smith, *J. Org. Chem.*, 1989, **54**, 4801–4807. See the experimental part in the ESI†.
- 24 J. L. Rainey and H. Adkins, *J. Am. Chem. Soc.*, 1939, **61**, 1104–1110.
- 25 In order to secure the optical activity of the intermediate **10c**, deprotection of the nitrogen atoms using trifluoroacetic acid at 0 °C in dichloromethane was performed. The <sup>1</sup>H NMR spectrum of the corresponding optically active product (**10d**; see the experimental part in the ESI†) was then recorded in the presence of the chiral shift reagent, europium (III) tris-(3-heptafluoropropylhydroxymethylene)-(+)-camphorate Eu(hfc)<sub>3</sub>, and it was compared to the spectrum obtained with a racemic sample. An ee value of 96% was deduced for compound **10c**.
- 26 N. K. Sharma and K. N. Ganesh, *Tetrahedron Lett.*, 2004, **45**, 1403–1406 and references cited therein.
- 27 We found that when the alkaline hydrolysis of the methyl esters was carried out in the first step, the pyroglutamic ring was opened, leading to the formation of the corresponding glutamic acid derivatives. This observation is consistent with some previous studies in the pyroglutamic acid series. See: A. G. Avent, H. M. E. Duggan and D. W. Young, *Org. Biomol. Chem.*, 2005, **3**, 2327–2332.